BOMBESIN ANALOGS FOR TREATMENT OF CANCERFIELD OF INVENTION

The present invention encompasses novel peptides that are antagonists to bombesin and bombesin like peptides and are useful in the treatment of cancer.

- 5 The invention particularly relates to the design and synthesis of the novel peptides incorporating α,α -amino acids in a site specific manner. The invention encompasses methods for the generation of these peptides, compositions containing the peptides and the pharmacological applications of these peptides especially in the treatment and prevention of cancer.

10 BACKGROUND OF THE INVENTION

Bombesin is a 14 amino acid peptide which was first isolated from the skin of the frog *Bombina bombina* (Anastasi et al., *Experientia*, 1971, 27, 166) and has the sequence:

- 15 pGlu-Gln-Arg-Leu-Gly-Asn-Gln-Trp-Ala-Val-Gly-His-Leu-Met-NH₂ (SEQ ID NO: 1)

Gastrin releasing peptide (GRP) is a 27 amino acid peptide isolated from the porcine gut. The last ten amino acids at the C-terminus of gastrin releasing peptide correspond with one amino acid alteration (3) to the last ten amino acids of bombesin, viz:

- 20 H-Gly-Asn-His-Trp-Ala-Val-Gly-His-Leu-Met-NH₂
(SEQ ID NO:2).

- It has been reported (J. H. Walsh and J. R. Reeve, *Peptides* 6, (3), 63-68, (1985)) that bombesin and bombesin-like peptides such as gastrin releasing peptide (GRP) are secreted by human small-cell lung cancer (SCLC) cells. It has
25 been postulated (P. J. Woll and E. Rozengurt, *PNAS* 85, 1859-1863, (1988)) that gastrin releasing factor antagonists would bind competitively to bombesin receptors in animals and would therefore be of use in the treatment of SCLC and/or in the control of clinical symptoms associated with this disease and due to hypersecretion of this peptide hormone. Analogues of bombesin / GRP have been shown to inhibit
30 the binding of gastrin releasing peptide to a SCLC cell line and to inhibit the growth of SCLC cells in-vitro and in-vivo (S. Mahmoud et al., *Cancer Research*, 1991, 51, 1798; Moody TW et al., *Life Sci.*, 1995, 56, 521; Moody TW et al.,

Peptides, 1996, 17, 1337). After Bombesin/GRP cell receptors were established on SCLC cells, receptors were also found to be present on human prostate cells. Reile H et al., (Prostate, 1994, 25: 29-38) showed that the PC-3 and DU-145 human prostate cancer cell lines possess specific high-affinity receptors for bombesin/GRP and are suitable models for the evaluation of anti-neoplastic activity of new bombesin/GRP antagonists in the treatment of androgen-dependent prostate cancer. Bombesin also increased the penetration of the two human prostatic carcinoma cell lines, the relatively indolent LNCaP cells and the aggressively growing and invasive PC-3 cells, in an in vitro invasion of reconstituted basement membrane (Matrigel) (Hoosein NM et al., J Urol, 149(5): 1209-1213). High-affinity binding sites for GRP were found on human colorectal cancer tissue (Preston, SR. et al, Br. J. Can., 1995, 71, 1087), suggesting that bombesin-like peptides may have a role in the pathogenesis of colorectal cancer, and bombesin receptor antagonists may be of value in the treatment of receptor-positive tumours. Inhibitory effects of bombesin/GRP antagonist RC-3095 and somatostatin analogue RC-160 were also seen on growth of HT-29 human colon cancer xenografts in nude mice (Radulovic S et al., Acta Oncol, 1994, 33(6): 693-701).

Studies with the anti-bombesin/GRP antibodies lead to the hypothesis that it may be possible to disrupt the autocrine growth cycle of bombesin/GRP using designed peptide receptor antagonists. Since then several types of Bombesin antagonists have been reported. These antagonists have been defined by type and position of the substitutions of the natural sequence. Early receptor antagonists suffered from low potency, lack of specificity, and toxicity, which presented serious problems with their scientific and therapeutic use.

More recent work has concentrated on modification of the carboxy terminal (C-terminal) region of these peptides to interrupt the receptor interaction utilizing a variety of different types of C-terminal modified analogs. These have included incorporation of D-amino acids, non-peptide bonds for example (psi. CH₂NH), amide, and ester modifications. These alterations gave rise to certain peptides having improved characteristics (Staley J et al., Peptides, 1991, 12(1): 145-9; Coy DH et al., J Natl Cancer Inst Monogr, 1992, 13: 133-9). Other patents that describes bombesin and related analogs are:

USP5,834,433 (1998)

USP 5,723,578 (1998)

USP 5,620,959 (1997)

USP 5,620,955 (1997)

5 USP 5,428,019 (1995)

USP 5,369,094 (1994)

USP 5,084,555 (1992)

A Bombesin/GRP antagonist (RC-3940-II) was found to inhibit the proliferation of SW-1990 human pancreatic adenocarcinoma cells in vivo and in
10 vitro (Qin, Y. et al., 1995, Int. J. Cancer, 63, 257). Similar effect was seen with bombesin/GRP antagonist RC-3095 on the growth of CFPAC-1 human pancreatic cancer cells transplanted to nude mice or cultured in vitro (Qin Y et al., Can Res, 1994, 54(4): 1035-41).

As reported earlier, the autocrine growth cycle of bombesin/GRP in
15 SCLC can be disrupted by bombesin/GRP antagonists such as [Psi 13,14] bombesin. Several bombesin analogues were solid phase synthesized and incubated with intact SCLC cells at 37°C in RPMI medium in a time course fashion (0-1080 minutes) to determine enzymatic stability. The proteolytic stability of the compounds was determined by subsequent HPLC analysis. [Psi 13, 14] Bombesin was found to be
20 very stable to metabolic enzymes (T_{1/2} = 646 min.) and inhibited SCLC xenograft formation in vivo in a dose-dependent manner (Davis TP et al., Peptides, 1992, 13(2): 401-7).

Female athymic nude mice bearing xenografts of the MCF-7 MII human breast cancer cell line were treated for 7 weeks with bombesin/GRP
25 antagonist (DTpi6, Leu13 psi[CH₂NH]-Leu14) bombesin (6-14)(RC-3095) injected subcutaneously daily at a dose of 20 µg and LHRH antagonist SB-75 (Cetrorelix) administered biweekly in the form of microgranules releasing 45 µg/ day. After 2 weeks of treatment, a significant inhibition of tumor volume was observed in the groups treated with RC-3095 alone or in combination with SB-75 (Yano T et al.,
30 Cancer, 1994, 73(4): 1229-38).

Pinski J et al., (Int. J. Cancer, 1994, 57(4): 574-580), demonstrated for the first time that the growth of gastrin-responsive human gastric carcinoma

MKN45 cell line xenografts in nude mice could be inhibited not only by somatostatin analogues, but also by administration of modern bombesin/GRP antagonists, such as RC-3095, or a combination of these. RC-3095 also effectively inhibited tumor growth in nude mice bearing xenografts of the human gastric cancer cell line Hs746T (Qin Y et al., J Cancer Res Clin Oncol, 1994,120(9):519-528).

This invention describes the preparation and use of peptide analogs of bombesin/GRP using constrained amino acids and their use for cancer therapy, alone, or in combination or as an adjunct to or with other chemotherapeutic agents and compounds.

The design of conformationally constrained bioactive peptide derivatives has been one of the widely used approaches for the development of peptide-based therapeutic agents. Non-standard amino acids with strong conformational preferences may be used to direct the course of polypeptide chain folding, by imposing local stereochemical constraints, in de novo approaches to peptide design. The conformational characteristics of α,α -dialkylated amino acids have been well studied. The incorporation of these amino acids restricts the rotation of ϕ , Ψ angles, within the molecule, thereby stabilizing a desired peptide conformation. The prototypic member of α,α -dialkylated amino acids, α -aminoisobutyric acid (Aib) or α,α -dimethylglycine has been shown to induce (β -turn or helical conformation when incorporated in a peptide sequence (Prasad and Balaram, (1984); CRC Crit. Rev. Biochem. 16, 307-347; Karle and Balaram (1990) Biochemistry 29, 6747-6756). The conformational properties of the higher homologs of α,α -dialkylated amino acids such as diethylglycine (Deg), di-n-propylglycine (Dpg) and di-n-butylglycine (Dbg) as well as the cyclic side chain analogs of α,α -dialkylated amino acids such as 1-aminocyclopentane carboxylic acid (Ac5c), 1-aminocyclohexane carboxylic acid (Ac6c), 1-aminocycloheptane carboxylic acid (Ac7c) and 1-aminocyclooctane carboxylic acid (Ac8c) have also been shown to induce folded conformation (Prasad et al., (1995), Biopolymers 35, 11-20; Karle et al., (1995); J. Amer. Chem. Soc. 117, 9632-9637). α,α -dialkylated amino acids have been used in the design of highly potent chemotactic peptide analogs (Prasad et al., (1996) Int. J. Peptide Proteins RCS. 48, 312-318).

The present invention exploits the conformational properties of α,α -

dialkylated amino acids for the design of biologically active peptide derivatives, taking bombesin as the model system under consideration. Furthermore, it has been shown that lipophilization of bioactive peptides improves their stability, bioavailability and the ability to permeate biomembranes (Dasgupta, P et al; 1999, Pharmaceutical Res. 16, 1047-1053; Gozes, I, et al 1996, Proc. Natl. Acad. Sci. USA, 93, 427-432). In the present invention, we have also synthesized peptide derivatives having N-terminal alkanoyl groups from C2-C16 carbon atoms, which retain anticancer activity.

The present invention exploits the conformational properties of α,α -dialkylated amino acids for the design of biologically active peptide derivatives, taking bombesin as the model system under consideration. Furthermore, it has been shown that lipophilization of bioactive peptides improves their stability, bioavailability and the ability to permeate biomembranes (Dasgupta, P et al; 1999, Pharmaceutical Res. 16, 1047-1053; Goes, L, et al., 1996, Proc. Natl. Acad. Sci. USA, 93, 427-432).

Throughout the specification and claims the amino acid residues are designated by their standard abbreviations. Amino acids denote L-configuration unless otherwise indicated by D or DL appearing before the symbol and separated from it by a hyphen. Throughout the specification and claims, the following abbreviations are used with the following meanings:

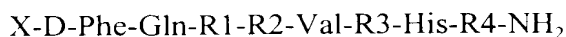
	BOP:	Benzotriazole-1-yl-oxy-tris-(dimethylamino)-phosphonium hexafluorophosphate
	PyBOP:	Benzotriazole-1-yl-oxy-tris-pyrrolidino-phosphonium hexafluorophosphate
25	TBTU:	2-(1H-Benzotriazole-1-yl)-1,1,3,3-tetramethyluronium tetrafluoroborate
	HBTU:	O-Benzotriazole-N,N,N',N'-tetramethyl-uronium-hexafluoro-phosphate
	HOBT:	1-Hydroxy Benzotriazole
30	DCC:	Dicyclohexyl carbodiimide
	DIPCDI:	Diisopropyl carbodiimide
	DIEA:	Diisopropyl ethylamine

DMF: Dimethyl formamide
DCM: Dichloromethane
NMP: N-Methyl-2-pyrrolidinone
TFA: trifluoroacetic acid

5

SUMMARY OF INVENTION

The present invention provides novel polypeptides of the following general formula,



wherein X is acetyl or straight, branched, or cyclic alkanoyl group from 3-16 carbon atoms, or X is deleted,

10 R1 is Trp or D-Trp,
R2 is Ala, Aib or Deg,
R3 is Gly, Aib, Deg, Dpg or Ac5c,
R4 is Leu or Ile or a hydrolyzable carboxy protecting group;
15 or a pharmaceutically acceptable salt of the polypeptide. At least one of R2 or R3 is a non-standard amino acid. The invention also encompasses methods for making the peptides, compositions containing the peptides and use of the peptides.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides novel polypeptides of the following general formula,

20 X-D-Phe-Gln-R1-R2-Val-R3-His-R4-NH₂
wherein X is acetyl or straight, branched, or cyclic alkanoyl group from 3-16 carbon atoms, or X is deleted,

25 R1 is Trp or D-Trp,
R2 is Ala, Aib or Deg,
R3 is Gly, Aib, Deg, Dpg or Ac5c,
R4 is Leu or Ile or a hydrolyzable carboxy protecting group;
or a pharmaceutically acceptable salt of the polypeptide. At least one of R2 or R3 is a non-standard amino acid.

30 A hydrolyzable carboxy protecting group are those groups which on hydrolysis converts to carboxylic group such as -COONH₂, -COOMe, etc.

The preferred alkanoyl groups are acetyl, n-butanoyl, n-hexanoyl, n-

octanoyl, lauroyl, myristoyl, palmitoyl, isohexanoyl, cyclohexanoyl, cyclopentyl-carbonyl, n-heptanoyl, n-decanoyl, n-undecanoyl and 3,7-dimethyloctanoyl.

Salts encompassed within the term "pharmaceutically acceptable salts" refer to non-toxic salts of the compounds of this invention. Representative salts and esters include:

5 acetate, ascorbate, benzenesulfonate, benzoate, bicarbonate, bisulfate, bitartrate, borate, camsylate, carbonate, citrate, dihydrochloride, methanesulfonate, ethanesulfonate, p-toluenesulfonate, cyclohexylsulfamate, quinate, edetate, edisylate, estolate, esylate, fumaxate, gluconate, glutamate, glycerophosphates, hydrobromide, 5 hydrochloride, hydroxynaphthoate, lactate, lactobionate, laurate, malate, maleate, 10 mandelate, mesylate, mucate, napsylate, nitrate, n-methylglucamine, oleate, oxalate, palmoates, pamoate (embonate), palmitate, pantothenate, perchlorates, phosphate/diphosphate, polygalacturonate, salicylates, stearate, succinates, sulfate, sulfamate, subacetate, succinate, tannate, tartrate, trifluoroacetate, tosylate and 15 valerate.

Other salts include Ca, Li, Mg, Na and K salts; salts of amino acids such lysine or arginine; guanidine, diethanolamine or choline; ammonium, substituted ammonium salts or aluminum salts.

The salts can be prepared by standard techniques.

20 Preferred peptides of this invention are:

D-Phe-Gln-Trp-Ala-Val-Aib-His-Leu-NH₂ (SEQ ID NO:3)

D-Phe-Gln-Trp-Aib-Val-Gly -His-Leu-NH₂ (SEQ ID NO:4)

D-Phe-Gln-D-Trp-Ala-Val-Aib-His-Leu-NH₂ (SEQ ID NO:5)

D-Phe-Gln-Trp-Aib-Val-Gly-His-Ile-NH₂ (SEQ ID NO:6)

25 D-Phe-Gln-Trp-Ala-Val-Aib-His-Ile-NH₂ (SEQ ID NO:7)

D-Phe-Gln-D-Trp-Ala-Val-Dpg-His-Leu-NH₂ (SEQ ID NO:8)

D-Phe-Gln-Trp-Deg-Val-Gly-His-Leu-NH₂ (SEQ ID NO:9)

D-Phe-Gln-Trp-Ala-Val-Ac5c-His-Leu-NH₂ (SEQ ID NO: 10)

Butanoyl-D-Phe-Gln-Trp-Ala-Val-Aib-His-Leu-NH₂ (SEQ ID NO:

30 11)

Octanoyl-D-Phe-Gln-Trp-Ala-Val-Aib-His-Leu-NH₂ (SEQ ID NO:

12)

The present invention also envisages methods of prevention and treatment of cancer using the polypeptides of the present invention, pharmaceutical compositions comprising such polypeptides and processes for their preparation. These peptides possess antagonist properties against bombesin and bombesin-like peptides and are useful in the prevention and treatment of malignant diseases.

5 Suitable routes for administration of the peptides are those known in the art and include oral, rectal, transdermal, vaginal, transmucosal, or intestinal administration; parenteral delivery, including intramuscular, subcutaneous, intraductal injections, as well as intrathecal, direct intraventricular, intravenous, 10 intraperitoneal, intranasal, or intraocular injections.

 Pharmaceutical compositions suitable for use in present invention include compositions wherein the active ingredients are contained in an effective amount to achieve its intended purpose. In addition to the active ingredients, these pharmaceutical compositions may contain suitable pharmaceutically acceptable 15 carriers, excipients, diluents, solvents, flavorings, colorants etc. The preparations may be formulated in any form including but not limited to tablets, dragees, capsules, powders, syrups, suspensions, slurries, time released formulations, sustained release formulations, pills, granules, emulsions, patches, injections, solutions, liposomes or nanoparticles.

20 The exact formulation, route of administration and dosage can be chosen by the individual physician in view of the patient's condition.

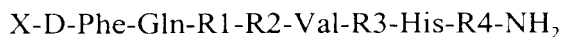
 The term "an effective amount" means that amount of a drug or pharmaceutical agent that will elicit the biological or medical response of a tissue, system, animal or human that is being sought.

25 Toxicity and therapeutic efficacy of the peptides of this invention can be determined by standard pharmaceutical procedures in cell cultures or experimental animals.

 The novel peptide analogs embodied in the present invention contain amino acids, namely α,α -dialkylated amino acids, which have been known to 30 induce highly specific constraints in the peptide backbone. The α,α -dialkylated amino acids, used in the present invention are synthesized from the corresponding ketones. In a preferred embodiment of the invention, the ketones are first converted

into the corresponding hydantoins which are hydrolyzed using a strong acid or base, preferably H_2SO_4 , HCl , NaOH or Na_2CO_3 to yield the aforesaid amino acids. In a preferred embodiment of the present invention, 60% sulphuric acid has been employed as the hydrolyzing agent.

5 The present invention also provides a solid phase synthesis process for the preparation of peptide analogs of the general formula (I):



wherein X is acetyl or straight, branched, or cyclic alkanoyl group from 3-16 carbon atoms or X is deleted,

10 R1 is Trp or D-Trp,
 R2 is Ala, Aib or Deg,
 R3 is Gly, Aib, Deg, Dpg or Ac5c,
 R4 is Leu or Ile

which comprises sequentially loading the corresponding protected α,α -dialkylated
15 amino acids in sequential cycles to the amino terminus of a solid phase resin, coupling the amino acids in the presence of conventional solvents and reagents to assemble a peptide-resin assembly, removing the protecting groups and cleaving the peptide from the resin to obtain a crude peptide analog.

 The novel peptides in the present invention have been generated by
20 using solid phase techniques or by a combination of solution phase procedures and solid phase techniques or by fragment condensation. These methods for the chemical synthesis of polypeptides are well known in the art (Stewart and Young, 1969, Solid Phase Peptide Synthesis, W.H. Freeman & Co.).

 In a preferred embodiment of the present invention the peptides were
25 synthesized using the Fmoc strategy, on a semi automatic peptide synthesizer (CS Bio. Model 536), using optimum side chain protection. The peptides were assembled from C-terminus to N-terminus. Peptides amidated at the carboxy-terminus were synthesized using the Rink Amide resin. The loading of the first Fmoc protected amino acid was achieved via an amide bond formation with the solid support,
30 mediated by Diisopropylcarbodiimide (DIPCDI) and HOBt. Substitution levels for automated synthesis were preferably between 0.2 and 0.6 mmole amino acid per gram resin.

The resin employed for the synthesis of carboxy-terminal amidated peptide analogs was 4-(2', 4'-Dimethoxyphenyl-Fmoc-aminomethyl)-phenoxymethyl derivatized polystyrene 1% divinylbenzene (Rink Amide) resin (100-200 mesh), procured from Calbiochem-Novabiochem Corp., La Jolla, U.S.A., (0.47 milliequivalent NH_2/g resin).

The N-terminal amino group was protected by 9-fluorenylmethoxycarbonyl (Fmoc) group. Trityl (trt) or t-butyloxycarbonyl (Boc) were the preferred protecting groups for imadazole group of Histidine residue. The hydroxyl groups of Serine, Threonine and Tyrosine were preferably protected by t-butyl group (tBu) 2,2,5,7,8-pentamethyl-chroman-6-sulfonyl (Pmc) or 2,2,4,7,-pentamethyl-dihydro-benzofuran-5 5-sulfonyl (Pbf) were the preferred protecting groups for the guandino group of Arginine. Trityl was the preferred protecting group for Asparagine and Glutamine and tertiary butyl group (tBu) was the preferred protecting group for Aspartic acid and Glutamic acid. The tryptophan residue was either left unprotected or used with Boc protection. The side chain amino group of Lysine was protected using Boc group preferably.

In a preferred embodiment of the invention, 2-8 equivalents of Fmoc protected amino acid per resin nitrogen equivalent were used. The activating reagents used for coupling amino acids to the resin, in solid phase peptide synthesis, are well known in the art. These include DCC, DIPCDI, DIEA, BOP, PyBOP, HBTU, TBTU, or HOBt. Preferably, DCC, DIPCDI/HOBt or HBTU/HOBT and DIEA were used as activating reagents in the coupling reactions.

The protected amino acids were either activated in situ or added in the form of preactivated esters known in the art such as NHS esters, Opfp esters etc. Atherton, E. et. al, 1988, J. Chem. Soc., Perkin Trans.I, 2887; Bodansky, M. in "The Peptides, Analysis, Synthesis and Biology (E. Gross, J. Meienhofer, eds) Vol. I, Academic Press, New York, 1979, 106.

The coupling reaction was carried out in DMF, DCM or NMP or a mixture of these solvents and was monitored by Kaiser test (Kaiser et al., Anal. Biochem., 34, 595-598 (1970)). In case of a positive Kaiser test, the appropriate amino acid was re-coupled using freshly prepared activated reagents.

After the assembly of the peptide was completed, the amino-terminal

Fmoc group was removed and then the peptide-resin was washed with methanol and dried. The peptides were then deprotected and cleaved from the resin support by treatment with trifluoroacetic acid, crystalline phenol, ethanedithiol, thioanisole and de-ionized water for 1.5 to 5 hours at room temperature. The crude peptide was
5 obtained by precipitation with cold dry ether, filtered, dissolved, and lyophilized.

The resulting crude peptide was purified by preparative high performance liquid chromatography (HPLC) using a LiChroCART® C,8 (250. Times. 10) reverse phase column (Merck, Darmstadt, Germany) on a Preparative HPLC system (Shimadzu Corporation, Japan) using a gradient of 0.1 % TFA in
10 acetonitrile and water. The eluted fractions were reanalyzed on Analytical HPLC system (Shimadzu Corporation, Japan) using a C18 LiChrospher®_g, WP-300 (300 X 4) reverse- phase column. Acetonitrile was evaporated and the fractions were lyophilized to obtain the pure peptide. The identity of each peptide was confirmed by electron-spray mass spectroscopy.

15 Synthesis Of Peptides

A peptide of the present invention can be made by exclusively solid phase techniques, by partial solid phase/solution phase techniques and/or fragment condensation. Preferred, semi-automated, stepwise solid phase methods for synthesis of peptides of the invention are provided in the examples discussed in the
20 subsequent section of this document.

The present invention will be further described in detail with reference to the following examples, as will be appreciated by a person skilled in the art are merely illustrative and should not be construed as limiting. Various other modifications of the invention will be possible without departing from the
25 spirit and scope of the present invention.

EXAMPLE 1

First loading on Rink Amide Resin

A typical preparation of the Fmoc-Leu-Rink Amide Resin was carried out using 0.5g of 4-(2',4'-Dimethoxyphenyl-Fmoc-aminomethyl)phenoxyethyl
30 derivatized polystyrene 1% divinylbenzene (Rink Amide) resin (0.7 mM/g) (100-200 mesh), procured from Advanced Chemtech, Louisville, KY, U.S.A., (0.7 milliequivalent NH₂ resin). Swelling of the resin was typically carried out in

dichloromethane measuring to volumes 10-40 ml/g resin. The resin was allowed to swell in methylene chloride (2 X 25 ml, for 10 min.). It was washed once in dimethylformamide (DMF) for 1 min. All solvents in the protocol were added in 20 ml portions per cycle. The Fmoc-protecting group on the resin was removed by following steps 3-7 in the protocol. The deprotection of the Fmoc group was checked by the presence of blue beads in Kaiser test. For loading of the first amino acid on the free amino (NH₂) group of the resin, the first amino acid, Fmoc-Leu-OH, was weighed in three to six fold excess, along with a similar fold excess of HOBt, in the amino acid vessel of the peptide synthesizer. These were dissolved in dimethylformamide (A.C.S. grade) (J.T.Baker, Phillipsburg, New Jersey, U.S.A.) and activated with DIPCDI, just prior to the addition to the resin in the reaction vessel of the peptide synthesizer. HOBt was added in all coupling reactions, especially in the case of Gln and His. The coupling reaction was carried out for a period ranging from 1-3 hours. The loading of the amino acid on the resin was confirmed by the presence of colorless beads in the Kaiser Test. The loading efficiency was ascertained by the increase of weight of the resin after the addition of the amino acid.

EXAMPLE 2

Synthesis of D-Phe-Gln-Trp-Ala-Val-Aib-His-Leu-NH₂ (SEQ ID NO: 3)

The synthesis of SEQ ID NO: 3, amidated at the carboxy- terminus, was initiated by using all of the resin loaded with Fmoc-Leu-OH as prepared in Example 1 above. This was subjected to stepwise deprotection and coupling steps as in steps 1-10 of the synthesis cycle. In each coupling reaction, a two to six fold excess of amino acid, DIPCDI and HOBt were used. Upon completion of synthesis and removal of the N-terminal Fmoc protecting group (steps 1-6 of the synthesis cycle), the peptide- resin was washed twice with methanol, dried and weighed to obtain 0.649g. This was subjected to cleavage in a cleavage mixture consisting of trifluoroacetic acid and scavengers, ethanedithol, crystalline phenol and thioanisole and water for a period of 1.5 to 5 hours at room temperature with continuous stirring. The peptide was precipitated using cold dry ether to obtain ~ 330 mg of crude peptide. The crude peptide was purified on a C18 preparative reverse phase HPLC column (250 X 10) on a gradient system comprising acetonitrile and water in

0.1 % TFA as described previously in the art. The prominent peaks were collected and lyophilized, reanalyzed on analytical HPLC and subjected to mass spectrometry. There was a good agreement between the observed molecular weight and calculated molecular weight (Calculated Mass ~ 983; Observed Mass ~ 984.2). The pure peptide was then used for bioassays.

EXAMPLE 3

Synthesis of D-Phe-Gln-Trp-Aib-Val-Gly-His-Leu-NH₂ (SEQ ID NO:4)

The synthesis, cleavage and lyophilization steps were carried out as in the Example 2 above using the appropriate amino acids. The calculated mass was ~ 969 and the observed mass was 970.4.

EXAMPLE 4

Synthesis of D-Phe-Gln-D-Trp-Ala-Val-Aib-His-Leu-NH₂ (SEQ ID NO:5)

The synthesis, cleavage and lyophilization steps were carried out as in the Example 2 above using the appropriate amino acids. The calculated mass was ~ 983 and the observed mass was 984.30.

EXAMPLE 5

Synthesis of D-Phe-Gln-Trp-Aib-Val-Gly-His-Ile-NH₂ (SEQ ID NO:6)

The synthesis, cleavage and lyophilization steps were carried out as in the Example 2 above using the appropriate amino acids. The calculated mass was ~ 969 and the observed mass was 970.2.

EXAMPLE 6

Synthesis of D-Phe-Gln-Trp-Ala-Val-Aib-His-Ile-NH₂ (SEQ ID NO:7)

The synthesis, cleavage and lyophilization steps were carried out as in the Example 2 above using the appropriate amino acids. The calculated mass was ~ 983 and the observed mass was 984.2.

EXAMPLE 7

Synthesis of D-Phe-Gln-D-Trp-Ala-Val-Dpg -His-Leu-NH₂ (SEQ ID NO:8)

The synthesis, cleavage and lyophilization steps were carried out as in the Example 2 above using the appropriate amino acids. The calculated mass was ~ 1039 and the observed mass was 1040.4.

EXAMPLE 8

Synthesis D-Phe-Gln-Trp-Deg-Val-Gly-His-Leu-NH₂ (SEQ ID NO:9)

The synthesis, cleavage and lyophilization steps were carried out as in the Example 2 above using the appropriate amino acids. The calculated mass was
5 ~ 997 and the observed mass was 998.5.

EXAMPLE 9

Synthesis of D-Phe-Gln-Trp-Ala-Val-Ac5c-His-Leu-NH₂ (SEQ ID NO: 10)

The synthesis, cleavage and lyophilization steps were carried out as in the Example 2 above using the appropriate amino acids. The calculated mass was
10 ~ 1009 and the observed mass was 1010.4.

EXAMPLE 10

Synthesis of Butanoyl-D-Phe-Gln-Trp-Ala-Val-Aib-His-Leu-NH₂ (SEQ ID NO: 11)

The conjugation of the butanoyl group at the N-terminal position was done on solid phase. The above peptide sequence was synthesized on resin as
15 described in Example 2. After the deprotection of D-Arg amino acid it was further coupled with butanoic acid in DMF using DIPCDI and HOBt. The cleavage and purification was further carried out following the standard protocol as described in Example 2. The final peptide was further analyzed by mass spectroscopy. The calculated mass and observed were in good agreement. (calculated mass = ~ 1053,
20 observed mass = 1054.2).

EXAMPLE 11

Synthesis of Octanoyl-D-Phe-Gln-Trp-Ala-Val-Aib-His-Leu-NH₂ (SEQ ID NO: 12)

The conjugation of the octanoyl group at the N-terminal position after the peptide synthesized as described in Example 2 was done on solid phase using
25 octanoic acid in DMF using DIPCDI and HOBt. The cleavage and purification was further carried out following the standard protocol as described in Example 2. The final purified peptide was further analyzed by mass spectroscopy. The calculated mass and observed were in good agreement. (calculated mass = ~ 1109, observed mass = 1110.5).

30

BIOLOGICAL ACTIVITY OF PEPTIDES

The cytotoxicity of the peptide analog was carried out by two day MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide] assay. MTT assay

is based on the principle of uptake of MTT, a tetrazolium salt, by metabolically active cells where it is metabolized by active mitochondria into a blue colored formazon product, which can be read spectrometrically (J. of Immunological Methods 65: 55-63, 1983). To prepare the MTT stock solution needed, MTT was dissolved in phosphate buffered saline with a pH of 7.4 to obtain an MTT concentration of 5 mg/ml; the resulting mixture was filtered through a 0.22 micron filter to sterilize and remove a small amount of insoluble residue. This filtered mixture was the MTT stock solution.

Briefly, for each tumor type, 10,000 cells were seeded in 96-well tissue culture plate and incubated with each peptide concentration individually in a CO₂ incubator for 48 hrs. The peptide analog at different concentrations was added once every 24 hrs during the incubation period. Control cultures, which were not treated with the peptide was similarly incubated. The assay was terminated by adding 100µg (20µl) of MTT to each well, incubating for three hours, decanting supernatant and finally adding 150 µl of dimethylsulphoxide to each well to dissolve the formazon. The plates were incubated for 15 minutes at 37°C and read spectrophotometrically at 540 nm; and cytotoxicity percentage was calculated by following formula:

$$\text{Cytotoxicity Percentage} = 100 \times [1 - X/R1],$$

where X= (absorbance of the treated sample at 540 nm-absorbance of a blank at 540 nm) and

R1 = (absorbance of the untreated control at 540nm) - (absorbance of the blank at 540nm).

Thus in each of the MTT cytotoxicity assay the percentage was calculated according to the above formula and was based on the proliferation of the untreated controls, the value of which was considered as 100%.

EXAMPLE 12

The biological activity of synthesized peptide SEQ ID NO:3 was tested on different human tumor cell lines such as HT-29 & PTC (colon), A549 (non small lung cell), KB (oral squamous cell), MCF7 & MDA.MB.453 (Breast), HuTu80 (duodenum), PA-1 (ovary), MOLT-4 (leukemia) and MIAPaCa2 (Pancreas) at various molar concentrations. The percentage cytotoxicity induced by different

concentrations of the peptide SEQ ID NO: 3 is summarized in the following table.

Cell Line	Percentage cytotoxicity at different concentrations					
	1 μ M	100n M	10 nM	1M	100pM	10pM
MCF 7	Nil	Nil	24.35 \pm 5	30.68 \pm 6	38.95 \pm 4.5	39.33 \pm 2.6
MIAPaCa2	33.3 \pm 4.5	30.3 \pm 4.2	33.2 \pm 6.7	36.4 \pm 0.5	28.2 \pm 4.5	27.4 \pm 4.5
HuTu80	12.2 \pm 4.0	15.5 \pm 4.7	14.3 \pm 3.5	13.3 \pm 4.0	14.7 \pm 4.2	10.3 \pm 3.5
KB	32.1 \pm 5.0	31.6 \pm 6.5	30.9 \pm 5.5	30.4 \pm 6.5	26.4 \pm 4.5	40.9 \pm 5.5
A549	30.7 \pm 6.5	23.6 \pm 4.5	32.2 \pm 5.5	32.4 \pm 4.5	25.2 \pm 3.5	30.5 \pm 3.5
HT29	25.4 \pm 5.5	17.8 \pm 4.5	11.8 \pm 5.0	20.3 \pm 4.5	19.9 \pm 5.5	18.7 \pm 4.5
PTC	17.9 \pm 2.5	27.7 \pm 2.8	27.7 \pm 3.6	23.8 \pm 2.8	26.5 \pm 3.8	80.0 \pm 7.1
MDA.MB.453	5.6 \pm 3.5	11.2 \pm 3.1	Nil	9.6 \pm 1.9	25.5 \pm 2.9	49.5 \pm 4.2
PA-1	31.2 \pm 5.1	34.2 \pm 5.8	25.4 \pm 4.2	36.1 \pm 6.1	40.1 \pm 6.2	37.7 \pm 3.9
MOLT-4	9.0 \pm 1.2	1.4 \pm 1.0	Nil	1.0 \pm 0.4	15.9 \pm 3.0	49.9 \pm 4.1

EXAMPLE: 13

The cytotoxic activity of other synthesized bombesin analogs was tested on eight human tumor cell lines namely HT-29, SW620, PTC (all colon), PA-1 (ovary), A549 (lung), HBL100 (breast), MOLT-4 (leukemia) and DU145 (prostate). The tumor cells were collected at exponential growth phase and resuspended in medium (1.5×10^6) cells/ml in RPMI 1640 containing 10% FBS). 150 μ l of medium was added to the wells of a 96-well tissue culture plate (Nunc, Denmark) followed by 30 μ l of cell suspension. The plate was left in incubator (37°C, 5% CO₂ overnight. 20 μ l of the peptide (10^{-7} x 10^{-10} M concentration) was added to marked wells of the 96-well plate. Each concentration was plated in triplicates. 20 μ l of medium alone was added to control wells while wells without cells served as blanks. A total volume of 200 μ l was ensured in each well and plate was left in incubator (37°C, 5% CO₂). After 72 hours of incubation an MTT assay was performed and percentage cytotoxicity was calculated with respect to control cells. Following tables show the cytotoxicity achieved on various cell lines at different concentrations.

PA-1

S.No	Percent Cytotoxicity			
	100 nM	10nM	1 nM	100 PM
SEQ ID:4	2.3 ± 2.9	4.3 ± 0.2	16.2 ± 2.9	12.6 ± 2.9
SEQ ID:5	8.8 ± 1.9	20.9 ± 5.3	16.0 ± 3.9	25.6 ± 6.3
SEQ ID:6	9.2 ± 1.0	8.7 ± 1.9	7.4 ± 1.0	11.1 ± 2.9
SEQ ID:7	9.6 ± 4.1	22.7 ± 3.4	25.6 ± 2.9	24.5 ± 4.2
SEQ ID:8	10.4 ± 3.7	20.4 ± 3.0	23.8 ± 4.2	23.3 ± 5.5

PTC

S.No	Percent Cytotoxicity			
	100 nM	10nM	1 nM	100 pM
SEQ ID:4	9.8 ± 1.7	2.1 ± 0.2	8.7 ± 1.5	14.9 ± 1.1
SEQ ID:5	20.4 ± 4.2	15.9 ± 2.4	23.0 ± 4.2	13.9 ± 2.2
SEQ ID:6	24.7 ± 5.2	10.4 ± 0.8	9.1 ± 0.7	10.1 ± 0.6
SEQ ID:7	9.3 ± 1.8	7.6 ± 0.7	12.4 ± 2.1	8.2 ± 0.9
SEQ ID:8	8.7 ± 2.1	5.4 ± 1.7	12.5 ± 1.7	12.3 ± 1.9

DU145

S.No	Percent Cytotoxicity			
	100 nM	10nM	1 nM	100 pM
SEQ ID:4	24.9 ± 3.2	23.4 ± 3.3	22.8 ± 4.1	23.2 ± 3.7
SEQ ID:5	32.3 ± 3.8	22.0 ± 3.4	10.6 ± 0.9	29.3 ± 2.9
SEQ ID:6	13.7 ± 0.9	16.6 ±	3.9 ± 5.2	12.1 ± 0.8
SEQ ID:7	NIL	NIL	ND	ND
SEQ ID:8	19.1 ± 2.1	22.5 ± 2-2	21.4 ± 6.2	28.1 ± 3.5

SW620

S.No.	Percent Cytotoxicity			
	100 nM	10 nM	1 nM	100 PM
SEQ ID: 4	34.3 ± 4.2	23.2 ± 2.0	27.8 ± 2.8	30.4 ± 3.2
SEQ ID: 5	25.6 ± 4.2	30.1 ± 4.0	29.7 ± 4.2	38.0 ± 3.8
SEQ ID: 6	23.5 ± 5.1	38.1 ± 7.3	33.5±5.2	24.8±4.2
SEQ ID: 7	25.4 ± 2.9	20.8 ± 1.9	32.0 ± 5.8	33.6 ± 5.8
SEQ ID: 8	29.4 ± 2.9	33.0 ± 3.8	20.6±3.9	20.6±3.9

HT29

S.No	Percent Cytotoxicity			
	100 nM	10nM	1 nM	100 PM
SEQ ID: 4	38.6 ± 5.3	38.9 ± 7.3	39.6 ± 4.3	43.3 ± 4.4
SEQ ID: 5	35.7 ± 2.8	44.4 ± 4.0	27.9 ± 2.9	42.0 ± 2.0
SEQ ID: 6	NIL	6.8 ± 0.7	26.7 ± 4.2	16.8 ± 0.5
SEQ ID: 7	15.5 ± 1.9	28.2 ± 2.8	ND	ND
SEQ ID: 8	34.8 ± 4.2	18.9 ± 4.2	34.7 ± 3.3	21.4 ± 3.1

MOLT4

S.No	Percent Cytotoxicity			
	100 nM	10nM	1 nM	100 PM
SEQ ID: 4	16.2 ± 0.6	28.7 ± 4.2	19.3 ± 1.8	28.5 ± 4.8
SEQ ID: 5	NIL	4.3 ± 0.6	6.4 ± 0.2	8.7 ± 0.6
SEQ ID: 6	NIL	20.4 ± 4.3	0.8 ± 0.1	11.0 ± 0.6
SEQ ID: 7	13.1 ± 0.3	NIL	NIL	ND
SEQ ID: 8	2.6 ± 0.1	12.8 ± 3.3	9.3 ± 0.2	16.6 ± 3.1

HBL

S.No	Percent Cytotoxicity			
	100 nM	10nM	1 nM	100 PM
SEQ ID: 4	25.0 ± 3.1	33.2 ± 5.2	30.6 ± 4.2	33.0 ± 3.6
SEQ ID: 5	19.4 ± 4.5	16.7 ± 3.6	31.6 ± 5.3	19.3 ± 2.7
SEQ ID: 6	17.0 ± 0.5	6.0 ± 0.4	1.2 ± 0.3	NIL
SEQ ID: 7	16.1 ± 3.9	7.0 ± 0.7	12.0 ± 0.7	4.0 ± 0.6
SEQ ID: 8	11.9 ± 2.1	14.4 ± 2.1	12.2 ± 1.9	12.1 ± 1.9

A549

S.No	Percent Cytotoxicity			
	100 nM	10nM	1 nM	100 PM
SEQ ID: 4	20.0 ± 2.2	20.6 ± 1.9	22.7 ± 2.9	20.7 ± 4.2
SEQ ID: 5	30.3 ± 4.3	22.2 ± 3.1	20.2 ± 4.2	25.2 ± 5.6
SEQ ID: 6	1.9 ± 0.6	3.2 ± 0.1	13.0 ± 0.8	12.4 ± 0.7
SEQ ID: 7	6.7 ± 2.0	17.9 ± 0.9	ND	ND
SEQ ID: 8	21.7 ± 3.3	20.7 ± 2.2	19.7 ± 3.1	17.0 ± 2.7

EXAMPLE 14

The cytotoxic effect of peptide sequences SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 11 and SEQ ID NO: 12, were studied by MTT assay which is based on the principle of uptake of MTT[3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl tetrazolium bromide], a tetrazolium salt by the metabolically active cells where it is metabolized by active mitochondria into a blue colored formazan product which can be read spectrophotometrically. Tumor cells KB (oral squamous), HuTu80 (Stomach), PTC and SW620 (colon), U87MG (Glioblastoma), HBL 100. (Breast), HeP2 (laryngeal) and L132 (Lung) were incubated with the peptide analogs for 48 hours at 37°C in a 96-well culture plate, followed by the addition of 100 µg MTT and further incubation of 1 hour. The formazan crystals formed inside the cells were dissolved with a detergent comprising 10% Sodium dodecyl sulfate and 0.01 N HCl and optical density read on a multiscan ELISA reader. The optical

density was directly proportional to the number of proliferating and metabolically active cells. Percent cytotoxicity of peptide analogs is shown in the following Table.

SEQ ID: 9

5

Cell	Percentage cytotoxicity at different concentrations					
	1 μ M	100n M	10 nM	1nM	100pM	10p M
KB	10.4 \pm 1.6	20.8 \pm 1.7	23.0 \pm 2.1	32.6 \pm 3.7	26.9 \pm 2.9	20.6 \pm 4.1
HuTu80	14.2 \pm 0.6	13.5 \pm 2.1	23.5 \pm 2.9	28.0 \pm 1.8	23.8 \pm 2.8	19.5 \pm 0.4
PTC	10.3 \pm 0.9	19.5 \pm 4.1	26.8 \pm 3.8	25.6 \pm 5.1	24.5 \pm 3.9	22.4 \pm 2.2
U87MG	10.0 \pm 0.0	21.4 \pm 0.1	20.0 \pm 0.0	21.8 \pm 0.1	11.9 \pm 4.1	0.0 \pm 0.0
SW620	21.6 \pm 2.1	25.8 \pm 2.8	33.2 \pm 2.9	30.8 \pm 0.6	28.9 \pm 0.2	15.1 \pm 0.3
HBL100	17.2 \pm 0.4	22.4 \pm 1.7	28.1 \pm 0.6	34.1 \pm 1.8	28.6 \pm 2.2	17.2 \pm 0.1
HeP2	21.6 \pm 1.8	17.8 \pm 0.3	28.5 \pm 3.1	21.3 \pm 2.2	14.6 \pm 0.6	0.0 \pm 0.0
L132	18.3 \pm 2.9	25.9 \pm 2.6	27.2 \pm 3.1	30.5 \pm 4.1	22.4 \pm 0.8	0.0 \pm 0.0

10

SEQ ID: 10

15

20

25

Cell	Percentage cytotoxicity at different concentrations					
	1 μ M	100n M	10 nM	1nM	100pM	10p M
KB	16.5 \pm 0.2	22.0 \pm 1.1	27.3 \pm 2.7	31.1 \pm 4.1	25.0 \pm 6.3	19.2 \pm 2.9
HuTu80	17.2 \pm 1.1	21.0 \pm 2.0	20.6 \pm 1.7	23.3 \pm 2.8	22.9 \pm 0.2	13.5 \pm 0.8
PTC	28.4 \pm 3.6	29.3 \pm 3.2	32.5 \pm 5.1	29.4 \pm 2.9	21.6 \pm 3.1	22.2 \pm 4.9
U87MG	10.0 \pm 0.0	15.0 \pm 0.5	20.0 \pm 0.0	25.6 \pm 2.1	16.5 \pm 0.5	11.6 \pm 1.7
SW620	22.2 \pm 2.1	19.4 \pm 1.8	25.5 \pm 2.8	22.4 \pm 1.7	20.9 \pm 0.6	16.7 \pm 0.2
HBL100	18.5 \pm 1.7	21.2 \pm 1.7	32.9 \pm 0.7	23.3 \pm 1.6	16.6 \pm 0.1	21.1 \pm 0.7
HeP2	19.9 \pm 1.5	26.3 \pm 1.7	27.5 \pm 2.8	27.2 \pm 2.6	19.1 \pm 0.6	1.7 \pm 0.1
L132	22.4 \pm 1.8	27.8 \pm 2.1	27.5 \pm 2.8	29.5 \pm 2.8	29.4 \pm 1.9	1.9 \pm 0.2

SEQ ID: 11

Cell	Percentage cytotoxicity at different concentrations					
	1 μ M	100n M	10 nM	1nM	100pM	10p M
KB	24.2 \pm 1.2	31.9 \pm 2.1	31.9 \pm 3.1	33.1 \pm 2.1	26.7 \pm 5.1	21.6 \pm 3.7
HuTu80	14.2 \pm 0.1	20.0 \pm 3.1	27.3 \pm 2.7	30.5 \pm 4.1	22.6 \pm 3.9	17.6 \pm 1.6
PTC	18.6 \pm 1.5	25.8 \pm 2.5	25.7 \pm 4.1	28.5 \pm 2.8	28.3 \pm 0.8	19.7 \pm 0.6
U87MG	1.0 \pm 0.1	15.5 \pm 0.6	20.0 \pm 0.0	24.2 \pm 1.7	26.5 \pm 2.6	21.9 \pm 2.1
SW620	23.7 \pm 1.4	21.0 \pm 1.5	31.5 \pm 2.6	35.1 \pm 2.2	25.9 \pm 3.8	20.4 \pm 0.3
HBL100	24.5 \pm 0.8	22.7 \pm 0.5	29.9 \pm 0.3	24.3 \pm 1.6	15.4 \pm 4.1	18.2 \pm 1.1
HeP2	21.9 \pm 2.1	23.9 \pm 1.1	34.6 \pm 2.2	37.1 \pm 3.3	20.1 \pm 0.0	15.1 \pm 0.3
L132	1.4 \pm 1.1	20.4 \pm 1.5	30.4 \pm 0.4	29.4 \pm 0.4	18.3 \pm 0.9	0.5 \pm 0.0

SEQ ID: 12

Cell	Percentage cytotoxicity at different concentrations					
	1 μ M	100n M	10 nM	1nM	100pM	10p M
KB	12.4 \pm 1.2	11.1 \pm 3.1	18.6 \pm 2.1	26.6 \pm 4.9	19.4 \pm 2.9	19.3 \pm 2.9
HuTu80	20.0 \pm 3.9	21.8 \pm 2.1	23.4 \pm 0.5	33.1 \pm 4.8	13.0 \pm 0.7	8.3 \pm 1.1
PTC	14.4 \pm 2.7	16.1 \pm 2.5	20.7 \pm 3.8	30.1 \pm 4.1	18.6 \pm 2.4	19.5 \pm 0.8
U87MG	15.4 \pm 3.1	13.1 \pm 2.3	27.5 \pm 2.9	28.3 \pm 1.9	22.1 \pm 3.8	13.1 \pm 2.2
SW620	22.6 \pm 1.1	25.3 \pm 0.6	36.1 \pm 1.9	32.2 \pm 2.6	38.4 \pm 2.8	34.8 \pm 0.4
HBL100	11.8 \pm 1.1	23.6 \pm 2.7	27.7 \pm 1.5	29.6 \pm 0.4	34.7 \pm 2.8	29.0 \pm 3.8
HeP2	28.7 \pm 0.8	25.6 \pm 0.4	29.2 \pm 1.1	28.9 \pm 0.5	24.4 \pm 0.1	10.0 \pm 0.0
L132	22.2 \pm 0.2	22.0 \pm 0.1	26.4 \pm 0.3	26.7 \pm 0.4	23.1 \pm 0.7	0.0 \pm 0.0

All publications referenced are incorporated by reference herein, including the nucleic acid sequences acid sequences and amino acid sequences listed in each publication. All the compounds and methods disclosed and referred to in the publications mentioned above are incorporated by reference herein, including those compounds disclosed and referred to in articles cited by the publications mentioned above.